Comparison of molecular weight distribution models for polymers characterized by g.p.c.

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A **semi-empirical model which employs polynomials based on general free radical polymerization kinetics, is developed to describe** the molecular **weight distribution data as well as to evaluate** the **average molecular weights of a variety of commercial thermoplastics including polystyrene,** poly(methyl methacrylate) and **low density polyethylene. These novel expressions are equally applicable** to a natural rubber sample with a bimodal distribution. Least-squares methods for the classical Schulz and Flory **distribution functions are introduced** to handle the g.p.c, data of the **above polymers. Comparison of** the **results collected from various analyses indicates** clearly that the **polynomial model is the most versatile one in the sense that it can be utilized to smooth out satisfactorily the molecular weight distribution data of many polymers. In general, the Wesslau distribution function is particularly good for** the highly **branched polyolefin and** the Schulz **model is fairly effective for** the addition **polymers of moderately sharp molecular weight distribution, presumably** with *Mw/Mn=3.0.* However, the Flory and Tung **distributions are found to be rather inferior in the present studies.** On the **basis of the current findings, a new procedure is suggested to facilitate the computations of the true average molecular weights from** g.p.c, data directly.

(Keywords: molecular weight distribution; polynomial function; least-squares analysis; molecular weight averages; gel permeation chromatography)

INTRODUCTION

Since the inception of gel permeation chromatography (g.p.c.) in the early $60s^{1-3}$, it has been routinely used to determine the molecular weight distribution of high polymers in many laboratories. This method has been recently improved by the coupling of a low-angle laser light scattering photometer^{4,5}. In spite of the rapid development of this particular experimental technique, the amount of work reported on the mathematical description of the molecular weight distribution data from g.p.c., which is of theoretical and practical importance, is rather scarce. Nevertheless, several classical models have been commonly applied to deal with the data obtained from the conventional fractionation experiments^{6a, 7}. The Flory⁸ and Schulz⁹ models can be derived theoretically but the Wesslau^{10,11} and Tung¹² distribution functions were devised empirically to suit some specific systems. These models are defined by one or two parameters and often describe less adequately the observed molecular weight distribution of most polymers. Thus analytical expressions that contain more than two parameters are postulated to cope with these involved situations.

It has been reported that a polymer may be considered as a blend of several components with known distributions^{13,14}. As such its molecular weight distribution can be specified by a linear function of the simple models that fit the individual components. This analysis hinges on the accuracy in differentiating the cumulative distribution curves. A second established multiparameter distribution function rests on a series that involves either Laguerre polynomials¹⁵⁻¹⁷ or Hermite polynomials¹⁸. The coefficients of these polynomials are evaluated from the moments of distributions which in many cases are difficult to estimate accurately, particularly the higher moments. In this paper, we derive a semi-empirical polynomial model which can readily be applied to analyse distribution data directly by means of a non-linear regression procedure. The effectiveness of these novel functions is tested using g.p.c, data which are also interpreted by various simple models.

Molecular weight distribution of fi'ee radical polymerization

The free radical polymerization of a monomer would produce an addition polymer of final molecular weight distribution depending predominantly on the rates of the individual kinetic steps involved as well as the duration of reaction. It may be described by a general kinetic scheme which includes four distinct elementary processes, viz. initiation, propagation, termination (combination and disproportionation) and chain transfer (to a monomer and/or a chain transfer agent) with their respective rate constants designated as k_i , k_p , k_t (combination), k'_i (disproportionation), k_{tr} (transfer to monomer) and k'_{tr} (transfer to chain transfer agent). If the instantaneous concentrations of the two essential reactants; monomer and chain transfer agent are denoted by $[M]$ and $[S]$ respectively at time t, then the rate of formation of \overline{X} -mer^{6h} expressed by the first derivative of the concentration of Xmer $[P_x]$ with respect to time t, is given by,

$$
\frac{d[P_x]}{dt} = (\phi \theta \psi + \phi^2 \theta^2 k_1 X/2) \exp(-\theta X) \tag{1}
$$

where the variables ϕ , θ and ψ are independent of the number of monomer units incorporated into the inactive chain P_x , and are defined respectively by,

$$
\phi = [I/(k_1 + k_1')]^{1/2}
$$
\n(1a)
$$
L_l = (l+1)H_{l+1} - \theta_0 H_l; l = 1, 2, ..., n-1
$$
\n(7b)

$$
\theta = \frac{k_{\text{tr}}[\mathbf{M}] + k_{\text{tr}}'[\mathbf{S}] + [I'(k_{\text{t}} + k_{\text{t}})]^{1/2}}{k_{\text{p}}[\mathbf{M}]}
$$
 (1b)

$$
\psi = k_{\rm tr}[\mathbf{M}] + k'_{\rm tr}[\mathbf{S}] + k'_{\rm t}\phi \tag{1c}
$$

Hereafter, we use the symbols I' and X to stand for the rate of initiation and the number of repeating units in the X-mer respectively. Two main assumptions are made in formulating equation (1), these are the steady-state approximation applied to the radicals and the independence of the rate constants of the size of the reactive species.

By using the Taylor series expansion, we can express the variable θ in the following form

$$
\theta = \theta_0 + \sum_{i=1}^{m} \theta_i t^i \tag{2}
$$

where the first term on the right hand side is independent of t and the constant θ_i is related to the ith derivative of θ evaluated at $t=0$. Equation (1) now becomes,

$$
\frac{d[P_x]}{dt} = (\phi \theta \psi + \phi^2 \theta^2 k_t X/2) \exp(-\theta_0 X) \exp\left(-X \sum_{i=1}^m \theta_i t^i\right)
$$
\n(3)

By expanding the exponential function $exp(- X \sum_{i=1}^{m} \theta_i t^i)$ in $i =$ series, we obtain from the foregoing equation,

$$
\frac{d[P_x]}{dt} = (\phi \theta \psi + \sum_{l=1}^{n} F_l(t) X^l) \exp(-\theta_0 X) \tag{4}
$$

where the coefficients F_1s' are functions of t and are intimately related to the variables ϕ , θ , ψ and constants k_t and θ_i s'. Integration of equation (4) results in,

$$
[P_x] = (G(t) + \sum_{l=1}^{n} H_l(t)X^l) \exp(-\theta_0 X)
$$
 (5)

where

$$
G(t) = \int_{0}^{t} \phi \theta \psi dt
$$
 (5a)

$$
H_{I}(t) = \int_{0}^{t} F_{I}(t) \mathrm{d}t \tag{5b}
$$

It follows that the number distribution functions $N'(X)$ may be written as,

$$
N'(X) = \frac{\partial [P_x]}{\partial X} \tag{6}
$$

$$
= \left(E + \sum_{l=1}^{n} L_l X^l\right) \exp(-\theta_0 X) \tag{7}
$$

where

$$
E = H_1 - \theta_0 G \tag{7a}
$$

$$
L_i = (l+1)H_{i+1} - \theta_0 H_i; \ l = 1, 2, ..., n-1 \ (7b)
$$

$$
L_n = -\theta_0 H_n \tag{7c}
$$

Then the differential distribution function $W(X)$ is equal to a polynomial,

$$
W(X) = \left(\sum_{l=1}^{n} R_l X^l\right) \exp(-\theta_0 X) \tag{8}
$$

where

$$
R_1 = E \Bigg/ \int_{0}^{\infty} XN'(X) dX \tag{8a}
$$

$$
R_l = L_l / \int_0^\infty XN'(X) dX; l = 2,3,\ldots, n
$$
 (8b)

To convert equation (8) into a useful form, we proceed to derive the integral distribution function,

$$
I(X) = \int_{0}^{X} W(X) \mathrm{d}X \tag{9}
$$

$$
= K - \left(K + \sum_{l=1}^{n} A_l X^l \text{exp}(-\theta_0 X) \right) \tag{10}
$$

where

$$
K = \sum_{l=1}^{n} \frac{l! R_l}{\theta_0^{l+1}}
$$
 (10a)

$$
A_{l} = \frac{1}{\theta_{0}} \sum_{s=1}^{n} \frac{s! R_{s}}{l! \theta_{0}^{s-l}}
$$
 (10b)

Normalization of function $W(X)$ leads to,

$$
I(y) = 1 - \left(1 + y + \sum_{l=2}^{n} N_l y^l\right) \exp(-y)
$$
 (11)

where $y = \theta_0 X$

$$
N_{l} = \sum_{j=1}^{n} \left(\frac{A_{j} - (j+1)A_{j+1}}{K} \right) \frac{j!}{l!}
$$
 (11b)

(lla)

$$
A_{1+n} = 0 \tag{11c}
$$

Consequently, equation (8) becomes,

$$
W(X) = \frac{\partial I(X)}{\partial X} \tag{12}
$$

$$
= \left\{ \theta_0 \sum_{l=1}^{n} \left[N_l - (l+1)N_{l+1} \right] y^l \right\} \exp(-y)
$$
\n(13)

with $N_1 = 1$ and $N_{1+n} = 0$. Eventually, the ath moment of

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the distribution $W(X)$ is equal to,

$$
U_a = \int\limits_0^\infty X^a W(X) \mathrm{d}X \tag{14}
$$

$$
= \theta_0^{-a} \sum_{l=1}^{n} \left[N_l - (l+1)N_{l+1} \right] \Gamma(1+a+l) \tag{15}
$$

where $\Gamma(1 + a + l)$ is a gamma function of the variable $(1 + a + l)$. Equations (11), (13) and (15) are used to analyse the molecular weight distribution data.

In practice, it is desirable to recast equation (11) to

$$
\{[1 - I(y)]\exp(y) - 1 - y\} = \sum_{l=2}^{n} N_l y^l
$$
 (16)

where the degree of the polynomial *n* should depend on circumstances. The value of the expression on the left hand side can be computed for each chain length X or experimental datum $I(y)$ provided that the constant θ_0 has been preset at a particular value. In the present analysis, the least-squares method is employed to determine the polynomial coefficients N_1s' . Together, we also compute the standard error of estimate of $I(y)$ on y defined by,

$$
\overline{SE} = \left\{ \frac{\sum_{y} \left[I(y) - 1 + \left(1 + y + \sum_{l=2}^{n} N_l y^l \right) \exp(-y) \right]^2}{N'' - 2} \right\}^{1/2} (17)
$$

where N'' is the total number of data points y or X. This computation cycle is repeated many times by varying the input value of θ_0 until a minimum *SE* is achieved.

Least-squares analyses for Schulz and other distribution models

The Schulz distribution function is one of the classical molecular weight distribution functions and is widely used particularly for vinyl polymerization terminated by coupling. This differential distribution function *W(M)* consists of two adjustable parameters α and b and is given as

$$
W(M) = \frac{(-\ln \alpha)^{b+2}}{\Gamma(b+2)} M^{b+1} \alpha^M
$$
 (18)

It describes the integral weight fraction of molecular weight M by an expression,

$$
I(M) = f(\alpha, b), g(\alpha, b) \tag{19}
$$

where

$$
f(\alpha, b) = \frac{(-\ln \alpha)^{b+2}}{\Gamma(b+2)}
$$
 (19a)

$$
g(\alpha, b) = \int_{0}^{M} M^{b+1} \alpha^{M} dM
$$
 (19b)

The integral function $g(x,b)$ may be conveniently evaluated by a numerical method if the values of α and b are known. Boyer¹⁹ and Mussa²⁰ have introduced two different graphical methods for determining the constants α and b. Here, we resort to a least-squares treatment for solving these two parameters.

A difference formula is first defined as,

$$
D = \sum_{M} [I(M) - f(\alpha, b) \cdot g(\alpha, b)]^2
$$
 (20)

of which a minimum value would offer the best-estimates of α and b designated respectively as α' and b'. This implies that

$$
F' = \left(\frac{\partial D}{\partial \alpha}\right)_x = 0\tag{21}
$$

$$
G' = \left(\frac{\partial D}{\partial b}\right)_{b'} = 0\tag{22}
$$

where

$$
F' = \sum_{M} \{ [I(M) - f(\alpha', b'), g(\alpha', b')] [f_x g(\alpha', b')+ f(\alpha', b'), g_x] \}
$$
 (21a)

$$
G' = \sum_{M} \{ [I(M) - f(\alpha', b'), g(\alpha', b')] [f_b \cdot g(\alpha', b')+ f(\alpha', b'), g_b \} \}
$$
(22a)

The various derivatives are,

$$
f_{x'} = \left(\frac{\partial f}{\partial \alpha}\right)_{x'} = \left(\frac{b' + 2}{\alpha' \ln \alpha'}\right) f(\alpha', b')
$$
 (23)

$$
f_{b'} = \left(\frac{\partial f}{\partial b}\right)_{b'} [\ln(-\ln \alpha') - \mu(b' + 1)] f(\alpha', b') \tag{24}
$$

$$
g_{\alpha} = \frac{1}{\alpha'} \int_{0}^{M} M^{b'+2} \alpha'^{M} dM
$$
 (25)

$$
g_{b'} = \int_{0}^{M} (\ln M) \cdot M^{b'+1} \alpha'^{M} dM
$$
 (26)

where $\mu(X)$ is a digamma function of X^{21} . The integrations in equation (25) and (26) are carried out numerically in this work. Equations (21) and (22) can be rearranged to obtain (respectively)

$$
\alpha' = \exp\bigg\{-\bigg[\frac{\Gamma(b'+2)\sum_{M} I(M)[f_x g(\alpha', b') + f(\alpha', b')g_x]}{\sum_{M} g(\alpha', b')[f_x g(\alpha', b') + f(\alpha', b'), g_x]}\bigg]^{1/2}\bigg\}
$$
\n(27)

$$
b' = \frac{1}{\ln(-\ln\alpha)}\ln\left[\frac{\Gamma(b'+2)\sum_{M}I(M)[f_{b'}g(\alpha',b')+f(\alpha',b')g_{b'}]}{\sum_{M}g(\alpha',b')[f_{b'}g(\alpha',b')+f(\alpha',b')g_{b'}]}\right] - 2
$$
\n(28)

An iterative method is applied to equations (27) and (28) to generate a series of approximate values of α' and β' . For each pair of these values, we also calculate the cor-

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responding function values of F' (equation (21a)), G' (equation $(22a)$) and S', where

$$
S' = F'^2 + G'^2 \tag{29}
$$

Practically, we select the best values of α' and b' as the pair that yields the smallest S' value. It will be illustrated that this is in fact an accurate and simple approach to evaluate the best-fit Schulz distribution curve. The standard error of estimate of $I(M)$ on M is equal to,

$$
\overline{SE} = [D/N'' - 2]^{1/2} \tag{30}
$$

where D is computed by equation (20) based on the finalselected values of α' and \dot{b}' , which also produce the a^{th} moment of *W(M),*

$$
U_a = \left(\frac{1}{-\ln \alpha'}\right)^a \frac{\Gamma(a+b'+2)}{\Gamma(a+b')} \tag{31}
$$

The Flory⁸ distribution function was first derived for the linear condensation polymers. In this case the differential distribution function for the X -mer is,

$$
W(X) = X(\ln P)^2 P^X \tag{32}
$$

where P is a probability constant. By analogy, we have the integral distribution function of the X-mer,

$$
I(X) = 1 + XP^X \ln P - P^X \tag{33}
$$

and a deviation function defined as,

$$
D = \sum_{X} (I(X) - 1 - XP^{X} \ln P + P^{X})^{2}
$$
 (34)

In the least-squares sense, the best value of P denoted by P' should satisfy the relationship,

$$
\left(\frac{\partial D}{\partial P}\right)_{P'} = 0\tag{35}
$$

An effective algorithm to a root of the foregoing equation is the Newton iteration method which obtains the successive approximations by,

$$
P'_{q} = P'_{q-1} - \frac{h(P'_{q-1})}{h'(P'_{q-1})}
$$
\n(36)

where the subscript q is for the qth repetition and the functions,

$$
h(P) = \sum_{X} [I(X) - 1 - XP^{X} \ln P + P^{X}] X^{2} P^{X}
$$
 (36a)

$$
h'(P) = \frac{\partial h(P)}{\partial P} = \sum_{X} (I(X) - 1 - XP^{X} \ln P + P^{X}) X^{3} P^{X-1}
$$

$$
- \sum_{X} X^{4} P^{2X-1} \ln P \qquad (36b)
$$

The ath moment of the distribution is,

$$
U_a = \Gamma(2+a)/(1-P')^a \tag{37}
$$

while the standard error of estimate of $I(X)$ on X may also be computed by equation (30) , but here D refers to equation (34).

Tung^{6a} has discussed data treatment using the Wesslau and Tung distribution models. The Wesslau distribution function contains two parameters M_0 and β ; the latter increases with the increase of the breadth of distribution. This unique feature is implicitly exhibited by the following equations:

$$
W(M) = \frac{1}{\beta \sqrt{\pi}} \frac{1}{M} \exp\left(-\frac{1}{\beta^2} \ln^2 \frac{M}{M_0}\right) \tag{38}
$$

$$
I(M) = \frac{1}{2} \left[1 + \text{erf}\left(\frac{1}{\beta} \ln \frac{M}{M_0}\right) \right]
$$
 (39)

$$
U_a = M^a_0 \exp(a^2 \beta^2 / 4) \tag{40}
$$

where erf (X) is an error function of X^{22} .

The Wesslau parameters β and M_0 are evaluated by finding the inverse transform of the error function using equation (39) and a relevant mathematical table²², i.e.

$$
\text{erf}^{-1}\left(\frac{1}{\beta}\ln\frac{M}{M_0}\right) = \frac{1}{\beta}\ln M - \frac{1}{\beta}\ln M_0 \tag{41}
$$

Hence a plot of $erf^{-1}(1/\beta \ln M/M_0)$ against $\ln M$ should produce a straight line whose gradient and intercept result in β and M_0 respectively. Analogously, we get,

$$
\overline{SE} = \left\{ \frac{\sum_{M} \left[I(M) - \frac{1}{2} - \frac{1}{2} \text{erf} \left(\frac{1}{\beta} \ln \frac{M}{M_0} \right) \right]^2}{N'' - 2} \right\}^{1/2}
$$
(42)

As far as the data analysis is concerned, the most advantageous model is perhaps the one postulated by Tung who has studied the molecular weight distribution of polyethylene using the equations listed below:

$$
W(M) = YZM^{Z-1}\exp(-YM^Z) \tag{43}
$$

$$
I(M) = 1 - \exp(-\,YM^Z) \tag{44}
$$

$$
U_a = Y^{-a/Z} \Gamma(1 + a/Z) \tag{45}
$$

where Y and Z are two adjustable parameters. Equation (44) can be readily rearranged to,

$$
\ln \ln [1 - I(M)]^{-1} = \ln Y + Z \ln M \tag{46}
$$

Once the parameters Y and Z are determined from the above linear plot, we can compute the

$$
\overline{SE} = \left\{ \frac{\sum \left[(M) - 1 + \exp(-YM^2) \right]^2}{N'' - 2} \right\}^{1/2}
$$
 (47)

This statistical parameter \overline{SE} is consistently⁴⁷ employed to indicate the goodness of fit of a particular integral distribution function to the observed cumulative weight fractions. Least-squares methods are applied to equations (41) and (46).

Figure 1 Plots of molecular weight distribution with $w \times 10$ against In M for sample S1. The data points are obtained from experiments and the curves **are predicted theoretically: Curve A;** polynomial model, curve B; Schulz model, curve C; Wesslau model, curve D; FIory model, and curve E; Tung model

Figure 2 (a) Plot of molecular weight distribution with $w \times 10$ against InM for sample S2. (b) Integral distribution curve with *I(M)* against InM for sample \$2. The data points are obtained from experiment and the curves are predicted by the polynomial model

EXPERIMENTAL

Six commercial thermoplastic samples and two biopolymer samples were studied altogether. These samples were designated as S1 and \$2 for polystyrene, M1 and M2 for poly(methyl methacrylate), E1 and E2 for low density polyethylene, and NI and N2 for natural rubber. Rudin *et al. 23* have reported the molecular weight distribution data of the same pair of poly(methyl methacrylate) samples (originally coded as samples A and C) recently. The g.p.c. data of the natural rubber samples (previously quoted as samples RRIM 703 and PB5/51) from different clones were acquired from the work of Subramanian²⁴.

Samples S1, \$2, E1 and E2 were analysed by the Polymer Supply and Characterization Centre of RAPRA, England. The g.p.c, columns which were calibrated by the polystyrene standards, were eluted with tetrahydrofuran at 25°C for polystyrene samples and with σ -dichlorobenzene plus 1% Ionox 330 as antioxidant at 138-140°C for the low density polyethylene samples.

Mathematical calculations were performed on a Univac 1100/11 computer system.

RESULTS AND DISCUSSION

The raw data are displayed in *Figures 1-5;* (plotting weight fraction, w, against logarithm of molecular weight M). Though the observed data points of the samples are not connected for the sake of simplicity, they all do obviously exhibit distinct unimodal distributions except sample N1 which is particularly chosen to represent the bimodal distribution. Moreover, sample N2 is conspicuously skewed to the high molecular weight region. In the ensuing analyses, various distribution functions will be applied to match these experimental data.

Figure 3 Plots of molecular weight distribution with wx 10 against InM for samples M1 and M2. The points are obtained from experiments and the curves are predicted by the polynomial model

Figure 4 Plots of molecular weight distribution with $w \times 10$ against InM for samples E1 and E2. The data points are obtained from experiments and the curves A and B are predicted by the polynomial and Wesslau models respectively

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It can be demonstrated that the constant $\theta_0 = 1/X_n$, where \bar{X}_n is the hypothetical number-average degree of polymerization at $t=0$. Hence the computation cycle using equations (16) and (17) is started with the first approximation of $\hat{\theta}_0$ taken as the reciprocal of \overline{X}_n . estimated by

$$
\bar{X}_n = \left[\bar{M}_0 \sum_{i=1}^{N''} w_i / M_i \right]^{-1}
$$
 (48)

where w_i is the weight fraction of the ith species of molecular weight M_i and \overline{M}_0 is the molecular weight of a

Figure 6 Plots of molecular weight distribution with w× 10 against InM for samples N1 and N2. The data points are obtained from experiments and the curves are predicted by the polynomial model

repeating unit. The θ_0 values is then successively decreased or increased until a smallest *SE* is produced. In any case the degree of n may be effectively kept below 10. The results obtained for all the samples are shown in *Table* 1.

Equations (27) and (28) are executed to create new sets of α' and b'. The first approximations of α' and b' are computed by,

$$
b' = \frac{2 - \Delta'}{1 - \Delta'}
$$
 (49)

$$
\alpha' = \exp\left[-\frac{(b'+2)}{\overline{M_{\rm w}'}}\right] \tag{50}
$$

where the polydispersity index,

$$
\Delta' = \overline{M'_{\rm w}/M'_{\rm n}}\tag{49a}
$$

Here, the apparent number-average and weight-average molecular weight are given respectively as:

$$
\overline{M'_n} = \left[\sum_{i=1}^{N''} w_i / M_i \right]^{-1} \tag{49b}
$$

$$
\overline{M'_{w}} = \sum_{i=1}^{N''} w_i M_i
$$
 (49c)

Figure 6 illustrates the variations of functions F' (equation (21a)) and G' (equation (22a)) with the iteration sequence \overline{N} for sample S1. Since the function F' is more responsive than the function G' to the parameter \overline{N} , the iterates that tend to vanish the former function are interpolated from the Figure. We have detected from the Figure that when \bar{N} increases from 12 to 13, the value of $ln \alpha'$ stays at -6.8694×10^{-6} , but the values of both parameters b' and

Figure 6 Iterative method applying to solve the Schulz parameters for sample S1; curve A: F' ×10⁻² against N and curve B: $G' \times 10$ against N

S' drop from -0.27889 to -0.27907 and from 266.0 to 46.6, respectively. Then appropriate combinations of α' and b' values are chosen within the above specified ranges and substituted into equations (27), (28) and (29). The smallest $S' = 0.0013$ decides the values of smallest $S' = 0.0013$ decides the values of $ln \alpha' = -6.8694 \times 10^{-6}$ and $b' = -0.27902$, which are considered as the best solutions to equations (21) and (22). These results are shown in *Table 1* together with those obtained by the same algorithm for other samples. It should be noted that the determining function S' is sensitive to at least the fourth decimal place of $\ln \alpha'$ and b'. Besides, this iterative procedure is relatively simple in that it does not require a good first approximation.

To compute the best value of the Flory parameter P' , we use equation (36). The calculation exercise is initiated by the first approximation $P' = 0.9990$, a value which makes the algorithms converge rapidly for all samples except N1 and N2, that do not yield convergent values at all. *Table 1* includes the lnP' values and also results obtained from the Wesslau (equation (41)) and Tung (equation (46)) models. The Tung distribution function does not hold for samples E2 and N1 because the parameter $Z < 1^{25}$.

Using the information provided in *Table 1,* equations (11), (19), (33), (39) and (44) would predict the cumulative weight fraction of any polymer species. The theoretical weight fraction w_i of an observed M_i is then readily computed by treating the function

2 I(X) or *I(M)* as a discrete variable, and hence we have $w_i = I(X_i) - I(X_{i-1}) = I(M_i) - I(M_{i-1}),$ where X_i is the number of repeating units of the species M_i . Some of these results are compared with those measured in *Figures 1-5. Figure 1* shows that the polynomial distribution function (equation (11)) is the most Ω suitable one for sample S1 and this coincides with the fact that it does give the smallest \overline{SE} value. In fact, these Figures are constructed to magnify the differences between the theoretical and experimental weight fractions over the whole molecular weight range covered in each case. *Figure 2(a)* confirms that such a plot is more informative than the conventional integral distribution -2 curve *(Figure 2(b))* which tends to compress the data points. Here the polynomial curve fits the observed data satisfactorily only when the molecular weight is neither too high nor too low, i.e. it holds over $11.0 < \ln M < 13.5$. This explains the enormous deviations between the predictions and measurements of the low- and high-order -4 average molecular weights for sample \$2 as will be touched on later. The Wesslau model is most compatible with sample E2 and so it is presented in *Figure 4.* Other curves in *Figures 3-5* are produced according to the polynomial distribution equation which fits these systems best.

In the present context, accordance between the es- ⁻⁶ timates and measurements has to be accomplished with respect to the average molecular weights which are based on a continuous frequency function, viz. the differential distribution function defined by equation (12). In terms of molecular weight M, we have

$$
W(M) = \frac{\partial I(M)}{\partial M}
$$
 (12a)

$$
U_a = \int\limits_0^\infty M^a W(M) dM \tag{14a}
$$

which correspond to equations (12) and (14) respectively. Equations (15), (31), (37), (40) and (45) are invoked to compute the various theoretical moments of distribution which in turn report the average molecular weights²⁶. To evaluate the average molecular weights from the experimental data, we estimate the derivative *W(M)* numerically. This is done by a least-squares fit of 3 adjacent data points to a parabola one at a time, i.e.

$$
I_i(M) = a_i + b_i \ln M + c_i(\ln M) \tag{51}
$$

where the empirical constants a_i , b_i and c_i are determined by the integral weight fractions $I(M_{i-1}), I(M_i)$ and $I(M_{i+1})$. These data points are arranged in the ascending order, i.e. the molecular weights $M_{i-1} < M_i < M_{i+1}$. It follows that, the experimental ath moment of distribution

$$
U_a = \sum_{i=2}^{N''} \int_{M_{i-1}}^{M_i} M^a W_i(M) dM
$$
 (52)

where $W_i(M)$ is the differential distribution function pertaining to the species M_i and it is equal to,

$$
W_i(M) = \frac{b_i}{M} + \frac{2c_i \ln M}{M} \tag{53}
$$

Figure 7 Differential distribution functional with $W(M) \times 10^6$ against InM for sample S1. The data points are derived from g.p.c, data and the curve is predicted by the polynomial model

As *Figure 7* is very similar to *Figure 1,* it illustrates that the $W(M)$ curve predicted by the polynomial model (equation (13)) is compatible with the data points calculated by equation (53) for sample SI. *Table 2* summarizes the values of the average molecular weights for all the samples. Obviously the polynomial model predictions are quite consistent with measurements for samples S1, M1, E1, M2 and N2, except that there are some noticeable discrepancies in the \overline{M}_n values for the last two samples, as is also shown in *Figure 3* and 5. In these two Figures, the theoretical curves do not conform to the experimental data at the low molecular weight ends. Although for sample \$2, considerable disagreement between some of the average molecular weight data from the polynomial model and experiment is encountered, as mentioned earlier, the values of \overline{M}_v and \overline{M}_w from the two sources are indeed in agreement.

The competency of this new model is also examined by a polyisoprene sample (N1) whose molecular weight distribution manifests two definite peaks at molecular weights around 1.5×10^5 and 2.3×10^6 *(Figure 5).* Surpris-

Table 2 Estimated average molecular weights based on continuous distribution

No.	Estimated Average Molecular Weight		Sample							
		Model	S ₁	S ₂	M1	M2	E1	E ₂	N ₁	N2
$\mathbf{1}$	$\overline{M_n}^3 \times 10^{-5}$	Expt.	1.244	0.756	0.097	0.481	0.105	0.232	1.581	4.134
		Polynomial	1.163	0.840	0.082	0.589	0.104	0.254	1.416	5.257
		Wesslau	1.209	0.718	0.100	0.383	0.111	0.268	1.980	4.114
		Schulz	1.050	0.548	0.065	0.359	0.069	0.067	0.209	3.810
		Flory	1.211	1,172	0.099	0.745	0.145	0.377		
		Tung	1,354	0.323	0.111	0.369	0.060			0.450
$\overline{2}$	\overline{M} $\sqrt{P} \times 10^{-5}$	Expt.	2.326	2.403	0.223	1.537	0.306	0.944	7.301	17.16
		Polynomial	2.308	2.318	0.222	1.544	0.308	0.847	6.967	17.00
		Wesslau	2.104	2.011	0.196	1.018	0.259	0.944	5.850	12.07
		Schulz	2.171	2.181	0.194	1.410	0.271	0.728	5.060	17.57
		Flory	2.140	2,071	0.175	1.317	0.256	0.666		$\qquad \qquad -$
		Tung	2.676	2.987	0.299	1.846	0.354			16.66
3	$M_{\rm m}{}^C$ x 10 ⁻⁵	Expt.	2.804	3.172	0.291	1.993	0.412	1.551	11.44	20.98
		Polynomial	2.762	2.922	0.293	1,989	0.410	1.170	10.54	20.51
		Wesslau	2.530	2.835	0.245	1.407	0.343	1.436	8.394	17.28
		Schulz	2.505	3.744	0.231	1.711	0.329	0.912	0.394	21,47
		Flory	2.422	2.344	0.198	1.490	0.290	0.754		
		Tung	3.005	3.688	0.346	2.225	0.431			21.04
4	\bar{M}_{2}^{d} x 10 ⁻⁵	Expt.	5.569	8.340	0.674	4.498	1.038	8.765	33.78	35.85
		Polynomial	5.216	5.906	0.701	4.447	0.982	2.996	27.12	34.07
		Wesslau	5.290	11.19	0.599	5.142	1.057	7.700	35.58	72.58
		Schulz	3.961	4.748	0.398	3.063	0.589	1.756	12.58	39.12
		Flory	3.633	3.516	0.297	2.235	0.435	1.130	-	
		Tung	4.288	6.807	0.538	3.856	0.761		<u></u>	41.22
5.	M_{Z+1} x 10 ⁻⁵ Expt.		9.249	16.69	1.034	7.053	1.762	19.01	49.26	47.36
		Polynomial	8.178	8.562	1.116	6.866	1.510	4.447	35.35	44.67
		Wesslau	11.08	44.19	1.463	18.79	3.260	4.129	150.8	304.9
		Schulz	5.417	6.849	0.564	4.414	0.848	2.601	18.77	56.80
		Flory	4.843	4.688	3.960	2.980	0.580	1.507	-	$\overline{}$
		Tung	5.391	9.805	0.707	5.377	1.072			61.18
6	$\overline{M}_{\mathsf{W}}/\overline{M}_{\mathsf{n}}$	Polynomial	2.37	3.48	3.55	3.38	3.95	5.36 f	7.44	3.90

aNumber-average molecular weight

bViscosity-average molecular weight under theta conditions

CWeight-average molecular weight

dZ-average molecular weight

e(Z+l)-average molecular weight

fValue *estimated* by Wesslau model

MWD models for g.p.c, characterized polymers: K. K. Chee

ingly, *Table 2* indicates that the values of \overline{M}_n , \overline{M}_v and \overline{M}_w can be estimated reasonably well by the polynomial method (equation (11)). The theoretical curve drawn in *Figure 5* is considered to be acceptable in view of the complicated feature of the distribution as well as the small SE=0.0119 *(Table 1)* obtained.

The Wesslau distribution function which best accounts for the g.p.c, data of sample E2 *(Figure 4),* furnishes comparable values of \bar{M}_{v} , \bar{M}_{w} and \bar{M}_{z+1} as quoted in *Table 2.*

The overall performance of the four simple models is also demonstrated by the *SE* data *(Table 1)* and average molecular weights *(Table 2).* As far as the curve-fitting of $I(M)$ data is concerned, the Schulz function is definitely superior to the other three for addition polymers with moderate polydispersity while the Wesslau function appears to be the most suitable one for polymers with broad molecular weight distribution, particularly polyolefins. When the data on average molecular weights are examined, it is evident that the Schulz and Wesslau distributions reproduce the observed low-order molecular weights \overline{M}_n , \overline{M}_v and \overline{M}_w , and high-order molecular weights \overline{M}_{w} , \overline{M}_{z} and \overline{M}_{z+1} reasonably well, respectively. In any event, the Flory and Tung distributions do not afford satisfactory predictions in the present investigations. These findings have some practical implications as explained below.

First of all, we will establish a distribution model which fits the g.p.c, data best by referring to the smallest value of *SE.* In this connection, it is not unreasonable to assume that the polynomial model is the choice as has been demonstrated in the majority of the cases. Under this specific condition, equation (16) is employed to deal with the experimental data. Now if the difference between the estimated and observed average molecular weights is within the experimental error which usually is not exceeding 10% at low molecular weights, then equation (15) will ultimately lead to the true values of various average molecular weights. This operation has an advantage in that it does not involve the approximate derivative $W(M)$, which is estimated either numerically (see equation (53)) or graphically. Unfortunately, both alogarithms are tedious and would inherently introduce considerable error in the final values of the average molecular weights. Thus, we have proposed a relatively simple approach to assess the average molecular weights accurately. Applications of this method to various commercial thermoplastics and a biomodal polymer have proved to be fruitful.

Incidentally, the values of absolute \overline{M}_n determined by osmometry, have been reported to be 0.193×10^5 and 1.10×10^5 for samples M1 and M2 respectively²³. These figures are substantially higher than those deduced from the g.p.c, measurements *(Table 2),* reflecting partially the approximate nature of the algorithm involved in estimating the derivative *W(M),* particularly in the low molecular weight region as also witnessed in *Figure 7* for sample S2. It has been commented that the overestimate from the osmometric data may be due to the experimental conditions that allow the low molecular weight fractions to pass through undesirably the membrane²

Table 2 also reports the values of the polydispersity index Δ . Recently, we have investigated the polydispersity effects on a particular property of polymer solutions²⁸. The present work justifies the use of Schulz distribution function for our purposes in those particular studies.

An immense amount of work has been done on the mathematical prediction of the molecular weight distribution in terms of the various kinetic parameters^{7,29,30}. However, to evaluate the relevant absolute rate constants from a set of a known molecular weight distribution data is a formidable task³¹. Preliminary investigations indicate that the present technique may be used to determine transfer constants via the coefficient θ_0 , which is inversely proportional to \overline{M}_n , as stated. Nevertheless, the relationships between the rate constants and the poynomial parameters N_t and θ_0 are too complicated to be dealt with even for a simple system such as the bulk polymerization of styrene initiated by 2,2'-azobisisobutyronitrile under controlled conditions. Furthermore, these rate constants are hardly mathematically separable from one another, except the one associated with the initiation step. This implies that the rate constant k_i , which can be determined rapidly from the polymerization kinetic data by a method proposed by us recently³², may also be derived from the molecular weight distribution data of the polymer.

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NOMENCLATURE

- *a* order of the moment U_a
- *ai* empirical constant associated with $I_i(M)$
- *b* Schulz parameter
- *b'* approximate value of b
- *bi* empirical constant associated with $I_i(M)$
- *ei* empirical constant associated with $I_i(M)$
- $f(\alpha,b)$ function defined by equation (19a)
- $f_{x'}$ function defined by equation (23)
- *f,,,* function defined by equation (24)
- $g(\alpha,b)$ function defined by equation (19b)
- $g_{\alpha'}$ function defined by equation (25)
- *gb"* function defined by equation (26)
- *h(P)* function defined by equation (36a)
- *h'(P)* function defined by equation (36b)
- *i J* summation index for power of t summation index
- $k_p^$ rate constant for propagation reaction
- *kt* rate constant for combination reaction
- *k~* rate constant for disproportionation reaction
- $k_{\rm tr}$ rate constant for transfer reaction to monomer
- k'_{tr} rate constant for transfer reaction to chain transfer agent
- *l* summation index or power of X
- *m* degree of Taylor series for θ
- *n* degree of various polynomials with argument X or Y
- q subscript for q^{th} repetition
- s summation index
- t time
- $w_i(w)$ weight fraction (normalized) of $M_i(M)$ or $X_i(X)$
- *Y* variable defined by equation (11a)
- A_l variable defined by equation (1b)
- *D* difference parameter
- *E* function defined by equation (7a)
- $F_l(t)$ complex function of t and other variables
- *F'* function defined by equation (21a)
- *G(t)* function defined by equation (5a)
- *G'* function defined by equation (22a)
- $H_t(t)$ function defined by equation (5b)
- *I'* initiation rate
- $I(M)$ integral distribution function of M
- *li(M)* empirical integral distribution function over a limited range of M around M_i
- $I(X)$ integral distribution function of X
- *I(y)* integral distribution function of ν
- *K* constant defined by equation (10a)
- L_{l} function defined by equation (7a)
- *L,* function defined by equation (7c)
- *M* molecular weight of polymer
- M_i molecular weight of i^{th} polymer species
- M_o Wesslau parameter
- \bar{M}_0 molecular weight of a repeating unit
- \bar{M}_n number-average molecular weight
- $\bar{M_{n}^{'}}$ apparent number-average molecular weight
- $\overline{M}_{\rm v}$ viscosity-average molecular weight under theta conditions
- $\bar{M}_{\rm w}$ weight-average molecular weight
- $\bar{M}'_{\rm w}$ apparent weight-average molecular weight
- z-average molecular weight \overline{M}_{z}
- \overline{M}_{z+1} $(z + 1)$ average molecular weight
- **[M]** instantaneous monomer concentration
- $N_{\rm r}$ variable defined by equation (11b)
- *N'(X)* number distribution function of X
- *N"* total number of g.p.c, data points
- \bar{N} integer indicating Nth interation
- *P* Flory parameter
- *p,* approximate value of P
- $[P_x]$ concentration of X-mer
- R_i function defined by equations (8a) and (8b)
- S' function defined by equation (29)
- *SE* standard error of estimate
- $[S]$ $U_a^$ instantaneous chain transfer agent concentration ath moment of distribution
-
- *W(M)* differential distribution function of M $W(X)$ differential distribution function of X
- $W_i(M)$ empirical differential distribution function over a limited range of M around M_i
- X number of repeating units of X -mer
- X_i number of repeating units in M_i
- \bar{X}_n hypothetical number-average degree of polymerization at $t = 0$
- Y Tung parameter
- Z Tung parameter
- Schulz parameter γ
- α' approximate value of α
- β Wesslau parameter
 θ X-independent parameter
- X -independent parameter defined by equation (1_b)
- θ_0 constant equal to θ at $t = 0$
- θ_i ith coefficient of Taylor series for θ
- $\mu(x)$ digamma function of x
- ϕ X-independent parameter defined by equation (1a) ψ X-independent parameter defined by equation (1c)
- $\Gamma(x)$ gamma function of x
- Δ polydispersity index

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